

# **Rungulla National Park Bush Blitz**

## ***Mammals & Birds***

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*Heather Janetzki & Dr Will Goulding*

Nomenclature and taxonomy used in this report is consistent with:

The Australian Faunal Directory (AFD)

<http://www.environment.gov.au/biodiversity/abrs/online-resources/fauna/afd/home>

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## List of contributors

List of contributors to this report.			
Name	Institution/affiliation	Qualifications/area of expertise	Level/form of contribution
<i>Heather Janetzki</i>	<i>Queensland Museum Biodiversity Section</i>	<i>Mammal/Bird Collection Manager</i>	<i>Mammal / Bird survey participant, identifications, report principal author mammals</i>
<i>Dr Will Goulding</i>	<i>Queensland Museum Biodiversity Section</i>	<i>Mammal/Bird Collection Manager</i>	<i>Mammal / Bird survey participant, identifications, report principal author birds</i>
<i>Dr Chris Burwell</i>	<i>Queensland Museum Biodiversity Section</i>	<i>Senior Curator, Entomology</i>	<i>Bird survey participant</i>

## Abstract

*Rungulla National Park had 14 species of mammals detected. It had good numbers of a small number of mammal species including Brushtail Possums. This exceptional landscape appeared well suited to Rock-wallabies with a population of Allied (*Petrogale assimilis*) close to the QPWS shed and several other macropod species. There are a few feral species which are likely to cause impacts on the abundance and suite of native mammal species present.*

*Rainy weather hampered trapping success particularly for microbats and smaller ground dwelling mammal species. An abundance of cane toads filling traps and hungry Brushtail Possums setting off multiple traps also reduced trapping success. As lack of detection does not provide proof of absence. Further mammal surveys at different times of the year would provide a more complete list.*

*73 species of birds were detected during the survey period in Rungulla National Park. Many of the same species were repeatedly encountered in different areas of the park and certain species that might be expected were not encountered. The mobility of many arid zone bird species makes it unlikely that some of these do not utilise habitat within the park at certain times or are in areas we did not visit. The surveys were conducted during the non-breeding period and we expect that further surveys at other times of the year, particularly during periods of more widespread flowering or seeding events, would reveal further species.*

*39 birds (14 species) were sampled for avian haemosporidians. Several new host parasite infections were identified, including the first infections in Rainbow Bee-eaters (*Merops ornatus*). These are awaiting molecular description.*

## 1. Introduction

Some mammal species had already been detected at Rungulla National Park, particularly the larger, more prominent kangaroos. Other macropods including *Onychogalea unguifera*, *Osphranter antilopinus* and *Aepyprymnus rufescens* had been sighted within 35 km of the QPWS shed. However, there was limited surveying of smaller mammals including dasyurids, rodents and microbats. Only one microbat was recorded from the area, *Taphozous troungtoni* at Fish Hole Creek. The landscape appeared to provide excellent habitat with multiple roosting sites and caves. *Pseudomys gracilicaudatus* had been collected 20 km SE of the QPWS shed on erosional escarpments and plains, so it and other native rodent species such as *Rattus* and *Zyomys* were targeted. There was also good tree coverage for arboreal species. Koalas had been previously recorded 10 km to the South and 30 km Northeast of the QPWS shed in the 1980's.

The birds of Rungulla National Park and surrounds have been relatively sparsely surveyed. Atlas of Living Australia has approximately 185 records of 56 bird species within the main park area and eBird Australia documents a similar suite of core bird species in nearby areas (e.g., Granite and Agate Creek areas, Robertson River crossing). Other notable nearby areas, such as Cobbold Gorge, have records of the same core bird species with only a few additional species, some of which might be expected with the larger permanent water bodies. The pre-survey expectations were therefore that we would find this core group of bird species, comprised of fairly common and easily observed species. However, we also expected to find further species that were noticeably absent from this core species group. In particular, we were hoping to encounter species of conservation interest such as Estrildid finch species historically known from close to the area, for example, Black-throated Finches *Peophila cincta cincta* (Ford 1986) or potentially Gouldian Finches *Erythrura gouldiae*.

The Einasleigh uplands forms an important biogeographic barrier, separating Cape York and northern Australia forms from southern races/subspecies in birds (e.g., Ford 1986), and is also

well placed near the edge of the Gulf Plains for eastern and western races/subspecies or distribution extents. Consequently, we were expecting some potentially interesting observations of closely related forms. Furthermore, we expected to find representatives of certain bird families not previously documented (e.g., Locustellids, Malurids, Neosittids, Pomatostomids, Climacterids to name a few), and further species of diverse under-represented groups that can be mobile across landscapes with temporally and spatially patchy resources (Estrildids, Cuculids, Meliphagids).

Avian malaria and related parasites (haemosporidians) that infect Australian native birds have been relatively understudied (See the online MalAvi database [<http://130.235.244.92/Malavi/>]). However, the available information from the region supports high avian host-specificity, particularly in natively occurring Haemoproteus parasites (e.g., Goulding et al. 2016, Clark et al. 2018). To our knowledge only a few avian haemosporidian studies have occurred in Queensland and all have been primarily coastal. Rungulla National Park sits in a previously unstudied inland location close to the meeting point of several of Queensland's bioregions but more specifically, close to the border of the Einasleigh Uplands and Gulf Plains. Consequently, we expected to identify novel haemosporidian parasites in previously unscreened hosts.

## 2. Methods

### 2.1 Site selection

The standard survey sites 1 and 2 were targeted as a priority. Standard site 1 looked promising with good grass coverage as cattle had been excluded from grazing the area.

Riparian vegetation accessible by foot (close to camp) and vehicle (Dead Horse Bend) were selected as initially, before the rains, waterholes would act as a drawcard for birds and mammals in the region. This too was the thought behind choosing Dutchman's Creek but also as this site included a change of vegetation with some *Triodia sp.* present. The second helicopter site was selected randomly with the Senior Ranger. It was hoped to return to a promising cave with evidence of microbats and an owl roost or another with Pandanus but this could not be located during the flight time. Rain impeded further site accessibility.

### 2.2 Survey techniques

Surveying for mammals included several trapping techniques.

Elliott trap transects targeted dasyurids and rodents; small wire cage traps for rodents (particularly *Rattus* species that won't go into enclosed Elliotts); larger wire traps and bandicoot traps for medium sized mammals. Traps were baited with peanut paste/ honey/ rolled oats or sausage meat and checked each morning. The transects were selected to cover a variety of vegetation types including riparian, rocky ridges, grassland.

Harp traps were set up across flyways for microbats. Unfortunately, the strings are easily detected by bats when wet and are not suitable for wet weather use, so they were removed from flooding areas and closed for several days of inclement weather.

Pitfall traps targeting smaller rodent and dasyurids were dug into the ground in a T formation with drift netting between. Fish traps were placed along the netting although these were to target reptiles and frogs, they can capture mammals. Shelter and moisture were provided in the bottom of the buckets. They were monitored regularly as meat ants could be problematic to the welfare of captures. Trap lids were closed when they were not able to be monitored or during wet weather.

A camera trap was set up in a likely area where rock-wallabies might traverse. The Senior Ranger, Nick Smith, also supplied several other camera traps for use during the last few days.

Diurnal birds were surveyed by walking through the landscape, making direct observations with binoculars and through call/vocalisation identification. Unless obvious, calls and vocalisations were further investigated to visually confirm species identification. A similar approach was employed during spotlighting for nocturnal birds. Vehicles were also used opportunistically to cover larger distances, during both the day and the night.

To survey birds for avian haemosporidian blood parasites, mist nets were set up along creek lines and areas offering sufficient vegetation cover. Playback of bird calls was used sparingly to draw in bird species to the netting area.

Less than ten mist nets were erected and opened during peak bird activity periods, primarily in the early morning when temperatures were appropriate. Mist nets were checked at least every 30 minutes. Methods for operating mist-nets followed Lowe (1989). Blood samples for parasite screening were collected following the commonly used venepuncture method (Owen 2011). The wing of each bird was gently extended in the standard Bander's holding position and a sterilised 26 - 28 gauge needle used to make a small puncture in the ulna vein on the underside of the wing. The exuding blood from this prick site was collected by capillary action into small microcapillary tubules (< 70 µl per bird). After blood collection, the prick site was checked for clotting (or pressure applied until bleeding ceased) and the bird immediately and quietly released in an unimpeded direction into habitat. A small droplet of blood from the microcapillary tube was used to create a blood smear (push-through method) to aid in visual identification of haemosporidian parasites, and the remaining blood sample was put into lysing buffer for storage and later use in molecular identification (PCR).

Spotlighting for birds and mammals using large spotlights was conducted from a vehicle along the main roads and on foot with hand-held and head torches around the sites.

Any signs of mammals and birds were also noted including feathers, skeletal remains, fur samples, diggings, scats and calls.

### **2.2.1 Methods used at standard survey sites**

Surveying at the sites occurred over a minimum of two days.

A transect of 25 Elliott traps was placed across the study site with the addition of two wire cage traps and two bandicoot traps. A harp trap was set up overnight for microbats. A set of 4 buckets and a drift net fence was dug for pitfall traps.

Each site was surveyed for at least an hour in the morning observing birds / recording calls / looking for signs of animals and again spotlighted at night for an hour. Birds flying over but not landing in the site were also noted. Flowering gums either in the standard sites and along the fence line were targeted, especially for gliders when spotlighting.

The conditions were not ideal due to the time of year (non-breeding and lower activity period) and the lack of widespread flowering resources.

### **2.3 Identifying the collections**

Mammals were identified through:- Observations (large macropods, arboreal species, possum trap captures) using morphological characteristics if not vouchered. Rock-wallabies from observations and photographs showing distinctive colouration and patterning such as cheek-stripe and with reference to the Queensland Museum specimen collection. Rodents on morphology and confirmed with skull characters. Microbats were keyed to species. (Churchill 2008, Van Dyck 2013) There are two difficult to separate *Scotorepens* species overlapping in this region. As the specimen collected was a female, and distinctive glans penis morphology not available, the skull characters were used to try and separated these. Gliders and possums identified using morphological features. (Cremona 2021)

All birds were identified from previous knowledge of the species and with reference to bird field guides (Pizzey & Knight 1999, Menkhorst et al. 2019). Avian blood smears were visually scanned for haemosporidian infections following Valkiunas (2005) and will be further screened using molecular methods (PCR) to complete formal identification.

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Queensland Museum Mammal Reference Collection.

### 3. Results and Discussion

Appendix 1 lists all mammals and birds recorded during the Bush Blitz. Collections made during this Bush Blitz will result in 5 mammal specimens, 7 mammal genetic samples and 39 bird blood samples being added to public collections (Queensland Museum research collections). Furthermore, 43 mammal records and 326 bird records (from 73 species) will be added to publicly accessible databases.

#### 3.1 Un-named or not formalised taxa

All mammal and bird species noted from Rungulla National Park are known species.

However, future research into the blood parasites from bird samples collected during the Bush Blitz survey may provide further clues to the distribution of these pathogens. Whilst molecular methods have yet to be applied to the blood samples collected, initial visual scans of the avian blood smears confirm several birds infected with Haemosporidians. This includes the first record of Haemoproteus infection in a Rainbow Bee-eater (*Merops ornatus*), which has potential to be a new species given the host-conservatism in these parasites (from a new location).

**Table 1. Putatively un-named or not formalised taxa**

Taxon	Comment
NIL	

#### 3.2 Putative new species (new to science)

In this report, 'putative new species' means an unnamed species that, as far as can be ascertained, was identified as a new species as a direct result of this Bush Blitz.

No putative new species were found during the survey for mammal and birds although there is a possibility a new species of a Haemoproteus infection of a Rainbow Bee-eater.

Table 2. Putative new species (new to science)	
Species	Comment
NIL	

### 3.3 Exotic and pest species

Table 3. Exotic and pest species recorded			
Exotic/pest species	Location sighted/observed	Indication of abundance	Comments
Rabbits	Adjacent to and ENE of QPWS shed, SS1	Frequently sighted	
Cattle	On road to Dead Horse Creek and evidence along Gilbert River.	Evident in more open country. Problematic particularly along water courses when low water levels.	Impact along river and grassy areas.
<i>Rattus rattus</i>	Ridge adjacent to QPWS shed	Multiple at site	While only located close to QPWS shed, problematic species
<i>Cane toads</i>	Along Gilbert River and surrounding area	Multiple at sites	Adults and toadpoles can impact native fauna. There was a high presence in vital water sources along the river prior to rains.

Feral species continue to have an impact. While the nature of the landscape appears to restrict accessibility of cattle to some areas and numbers have been greatly reduced, where they do occur, the pervasive effect of grazing is likely to impact seed-eating, ground foraging and nesting birds as well as rodents. Their trampling also reduces seedling recruitment and can cause erosion. Water degradation from dung was evident when the river levels were reduced to smaller water holes.

While the devastating impacts on island and the urban/ bush interface fauna is well known, the impact of Black rats on native wildlife in more remote locations is less documented. They affect wildlife through competition, predation, and the spreading of disease. (Banks & Hughes, 2012; Banks & Smith 2015) These *Rattus rattus* are over 100 km from the next most easterly site. Just how far they have dispersed within the National Park is not known although they are highly mobile in areas of human interference (as evident with a Rungulla NP Black rat ending up in Canberra in a Bush Blitz container).

### 3.4 Threatened species

Table 4. Threatened species			
Species	Listing status and level (EBPC, State/Territory)	Location sighted/observed	Indication of abundance
NIL			

No mammal or bird species sighted at Rungulla National Park are on the threatened species list.

### 3.5 Range extensions

Table 5. Range extensions or significant infill in distribution records for species			
Species	Location sighted/observed	Distance from nearest known record (km)	Comments
NIL			

### 3.6 Genetic information

All vouchered specimens were tissue sampled and lodged with the Queensland Museum's molecular identities collection for future researchers to access. Similarly, bird blood samples from birds captured and released are also with this museum collection for future researcher access. Any publications utilising these samples will also have sequences lodged in publicly accessible databases such as GenBank (and MalAvi for Avian Haemosporidia).

## 4. Information on species lists

There were minimal surveys previously done within the reserve although there are some records for Rungulla National Park and adjoining regions further east on Wildnet and the Atlas of Living Australia, particularly those of the Northern Gulf Research Group.

Previously encountered bird species for Rungulla National Park largely came from Atlas of Living Australia (<https://biocache.ala.org.au>) and eBird Australia (<https://ebird.org/australia/>) for nearby areas (e.g., Granite and Agate Creek areas, Robertson River crossing).

## 5. Information for land managers

A number of wet weather issues restricted survey outcomes. These included bats being able to detect wet harp traps, site access issues with the significant flooding, a reduction in the number of multiple days possible at any site, an invasion of traps by cane toads and only a small area of the park surveyed. Consequently, it would be good if any unknown skulls / remains of any mammals/ birds found while working in the park could be sent to the Queensland Museum for identification for addition to the species list. In particular, the intricate cave and hollow features of this landscape provide important roosts for bats and owls (we heard mention of a cave with owl pellets). Should skeletons of microbats or owl pellets with skulls or prey remains be found in caves, these would be useful to collect to provide greater insight into the suite of smaller mammal inhabitants of the region. This might also give an indication if *Rattus rattus* has dispersed far from the QPWS shed.



## 6. Conclusions

The incredible landscape provides good refuge for macropod species particularly the Allied Rock-wallaby. There was an abundance of a few arboreal species including gliders and particularly the Common Brushtail Possum. Given the Northern Brushtail subspecies, *Trichosurus vulpecula arnhemensis* is endangered, it is worth monitoring the arid areas where the north Queensland subspecies, *T.v. eburacensis* occurs. Many microbats were sighted but the limited call references (for acoustic monitoring identification) for this region of Queensland, combined with the need to often have an animal in the hand to look closely at identifying characteristics, means more work on the microbat group in this area is needed.

The number of bird species encountered in the national park was expected and will no doubt grow with further observations at different times of the year and with the varying context of the park within regional weather conditions.

## Acknowledgements

We wish to thank the Ewamian Traditional Owners (Ken Georgetown, Barry Fisher, Jimmy Richards) of the land for allowing access to this stunning landscape and for their local knowledge of the region; Nick Smith and the QPWS Rangers for their preparation of the camping area, local knowledge, providing access to research areas, digging pitfalls (Thanks Roy Mortensen and Luke Parnell) and providing extra camera traps to monitor wildlife; the Bush Blitz crew (Including Helen Cross, Courtney Webber) and chef (Robbie Bayliss), for their outstanding organisation and wonderful camp setup and support; helicopter pilot and personnel for the smooth travel (Clayton & Brett); BHP staff (Daniel Lachenicht & Kath Taske) volunteer teachers (Louise, Janet, Monica & Michael) and Earthwatch staff (Sabrina Trocini, Jock Mackenzie) for their assistance with trap setup and observations; James Cook University and Herbarium staff, particularly Matt Barrett for providing vehicle transport and assistance with gear; other participants for their freely given knowledge and great company.

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Appendix 1. List of mammals and birds recorded during the Rungulla Bush Blitz						
Family	Species	Common name	Putative new species	Threatened (EPBC Act)	Threatened (QLD Act)	Exotic/ pest
<b>Mammals</b>						
Trachyglossidae	<i>Tachyglossus aculeatus</i>	Echidna	No	No	No	No
Petauridae	<i>Petaurus notatus</i>	Kreff's Glider	No	No	No	No
Petauridae	<i>Petaurus norfolcensis</i>	Squirrel Glider	No	No	No	No
Phalangidae	<i>Trichosurus vulpecula</i>	Common Brushtail Possum	No	No	No	No
Macropodidae	<i>Macropus giganteus</i>	Eastern Grey Kangaroo	No	No	No	No
Macropodidae	<i>Osphranter robustus</i>	Common Wallaroo	No	No	No	No
Macropodidae	<i>Petrogale assimilis</i>	Allied Rock-wallaby	No	No	No	No
Macropodidae	<i>Wallabia bicolor</i>	Swamp Wallaby	No	No	No	No
Vespertilionidae	<i>Scotorepens sanborni</i>	Northern Broad-nosed bat	No	No	No	No
Vespertilionidae	<i>Vespadelus finlaysoni</i>	Finlayson's Cave Bat	No	No	No	No
Canidae	<i>Canis familiaris dingo</i>	Dingo	No	No	No	No
Muridae	<i>Rattus rattus</i>	Black Rat	No	No	No	Yes
Leporidae	<i>Oryctolagus cuniculus</i>	Rabbit	No	No	No	Yes
Bovidae	<i>Bos taurus</i>	Cattle	No	No	No	Yes
<b>Birds</b>						
Acanthizidae	<i>Gerygone olivacea</i>	White-throated Gerygone	No	No	No	No
Acanthizidae	<i>Smicrornis brevirostris</i>	Weebill	No	No	No	No
Accipitridae	<i>Accipiter fasciatus</i>	Brown Goshawk	No	No	No	No
Accipitridae	<i>Aquila audax</i>	Wedge-tailed Eagle	No	No	No	No
Accipitridae	<i>Haliastur sphenurus</i>	Whistling Kite	No	No	No	No
Accipitridae	<i>Haliaeetus leucogaster</i>	White-bellied Sea-eagle	No	No	No	No
Accipitridae	<i>Milvus migrans</i>	Black Kite	No	No	No	No
Aegothelidae	<i>Aegotheles cristatus</i>	Australian Owlette-nightjar	No	No	No	No
Alcedinidae	<i>Ceyx azureus</i>	Azure Kingfisher	No	No	No	No
Alcedinidae	<i>Dacelo leachii</i>	Blue-winged Kookaburra	No	No	No	No
Alcedinidae	<i>Dacelo novaeguineae</i>	Laughing Kookaburra	No	No	No	No
Alcedinidae	<i>Todiramphus sanctus</i>	Sacred Kingfisher	No	No	No	No

Family	Species	Common name	Putative new species	Threatened (EPBC Act)	Threatened (QLD Act)	Exotic/ pest
Anatidae	<i>Anas superciliosa</i>	Pacific Black Duck	No	No	No	No
Ardeidae	<i>Ardea pacifica</i>	White-necked Heron	No	No	No	No
Ardeidae	<i>Egretta intermedia</i>	Intermediate Egret	No	No	No	No
Ardeidae	<i>Egretta novaehollandiae</i>	White-faced Heron	No	No	No	No
Ardeidae	<i>Nycticorax caledonicus</i>	Nankeen Night-Heron	No	No	No	No
Artamidae	<i>Artamus minor</i>	Little Woodswallow	No	No	No	No
Artamidae	<i>Cracticus nigrogularis</i>	Pied Butcherbird	No	No	No	No
Artamidae	<i>Cracticus torquatus</i>	Grey Butcherbird	No	No	No	No
Artamidae	<i>Gymnorhina tibicen</i>	Australian Magpie	No	No	No	No
Artamidae	<i>Strepera graculina</i>	Pied Currawong	No	No	No	No
Burhinidae	<i>Burhinus grallarius</i>	Bush Stone-curlew	No	No	No	No
Cacatuidae	<i>Cacatua galerita</i>	Sulphur-crested Cockatoo	No	No	No	No
Cacatuidae	<i>Calyptorhynchus banksii</i>	Red-tailed Black Cockatoo	No	No	No	No
Cacatuidae	<i>Eolophus roseicapilla</i>	Galah	No	No	No	No
Campephagidae	<i>Coracina novaehollandiae</i>	Black-faced Cuckoo-shrike	No	No	No	No
Campephagidae	<i>Lalage tricolor</i>	White-winged Triller	No	No	No	No
Charadriidae	<i>Euseyornis melanops</i>	Black-fronted Dotterel	No	No	No	No
Charadriidae	<i>Vanellus miles</i>	Masked Lapwing	No	No	No	No
Ciconiidae	<i>Ephippiorhynchus asiaticus</i>	Black-necked Stork	No	No	No	No
Columbidae	<i>Geopelia cuneata</i>	Diamond Dove	No	No	No	No
Columbidae	<i>Geopelia placida</i>	Peaceful Dove	No	No	No	No
Columbidae	<i>Geophaps scripta peninsulae</i>	Squatter Pigeon	No	No	No	No
Columbidae	<i>Phaps chalcoptera</i>	Common Bronzewing	No	No	No	No
Corcoracidae	<i>Struthidea cinerea</i>	Apostlebird	No	No	No	No
Corvidae	<i>Corvus coronoides</i>	Australian Raven	No	No	No	No
Corvidae	<i>Corvus orru</i>	Torresian Crow	No	No	No	No
Cuculidae	<i>Centropus phasianinus</i>	Pheasant Coucal	No	No	No	No
Dicaeidae	<i>Dicaeum hirundinaceum</i>	Mistletoebird	No	No	No	No
Dicruridae	<i>Dicrurus bracteatus</i>	Spangled Drongo	No	No	No	No
Estrildidae	<i>Stizoptera bichenovii</i>	Double-barred Finch	No	No	No	No
Falconidae	<i>Falco berigora</i>	Brown Falcon	No	No	No	No
Falconidae	<i>Falco cenchroides</i>	Nankeen Kestrel	No	No	No	No

Family	Species	Common name	Putative new species	Threatened (EPBC Act)	Threatened (QLD Act)	Exotic/ pest
Gruidae	<i>Antigone antigone</i>	Sarus Crane	No	No	No	No
Hirundinidae	<i>Petrochelidon ariel</i>	Fairy Martin	No	No	No	No
Hirundinidae	<i>Petrochelidon nigricans</i>	Tree Martin	No	No	No	No
Megapodiidae	<i>Alectura lathami</i>	Australian Brush-turkey	No	No	No	No
Meliphagidae	<i>Conopophila rufogularis</i>	Rufous-throated Honeyeater	No	No	No	No
Meliphagidae	<i>Entomyzon cyanotis</i>	Blue-faced Honeyeater	No	No	No	No
Meliphagidae	<i>Gavicalis virescens</i>	Singing Honeyeater	No	No	No	No
Meliphagidae	<i>Lichmera indistincta</i>	Brown Honeyeater	No	No	No	No
Meliphagidae	<i>Melithreptus albogularis</i>	White-throated Honeyeater	No	No	No	No
Meliphagidae	<i>Philemon citreogularis</i>	Little Friarbird	No	No	No	No
Meliphagidae	<i>Philemon corniculatus</i>	Noisy Friarbird	No	No	No	No
Meliphagidae	<i>Stomiopera flava</i>	Yellow Honeyeater	No	No	No	No
Meropidae	<i>Merops ornatus</i>	Rainbow Bee-eater	No	No	No	No
Monarchidae	<i>Grallina cyanoleuca</i>	Magpie-lark	No	No	No	No
Monarchidae	<i>Myiagra rubecula</i>	Leaden Flycatcher	No	No	No	No
Oriolidae	<i>Oriolus sagittatus</i>	Olive-backed oriole	No	No	No	No
Pachycephalidae	<i>Pachycephala rufiventris</i>	Rufous Whistler	No	No	No	No
Pardalotidae	<i>Pardalotus striatus</i>	Striated Pardalote	No	No	No	No
Petroicidae	<i>Microeca fascinans</i>	Jacky Winter	No	No	No	No
Phasianidae	<i>Coturnix ypsilophora</i>	Brown Quail	No	No	No	No
Podargidae	<i>Podargus strigoides</i>	Tawny Frogmouth	No	No	No	No
Psittacidae	<i>Aprosmictus erythropterus</i>	Red-winged Parrot	No	No	No	No
Psittacidae	<i>Platycercus adscitus</i>	Pale-headed Rosella	No	No	No	No
Psittacidae	<i>Trichoglossus haematodus</i>	Rainbow Lorikeet	No	No	No	No
Ptilonorhynchidae	<i>Chlamydera nuchalis</i>	Great Bowerbird	No	No	No	No
Recurvirostridae	<i>Himantopus himantopus</i>	Black-winged Stilt	No	No	No	No
Rhipiduridae	<i>Rhipidura albiscapa</i>	Grey Fantail	No	No	No	No
Rhipiduridae	<i>Rhipidura leucophrys</i>	Willie Wagtail	No	No	No	No
Strigidae	<i>Ninox boobook</i>	Boobook Owl	No	No	No	No